

This Page Is Inserted by IFW Operations
and is not a part of the Official Record

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

**As rescanning documents *will not* correct images,
please do not report the images to the
Image Problems Mailbox.**

(19)



Europäisches Patentamt
European Patent Office
Office européen des brevets



(11) Publication number:

0 339 607 B1

(12)

EUROPEAN PATENT SPECIFICATION

(43) Date of publication of patent specification: 10.02.93 (51) Int. Cl.⁵: **A61K 35/28, A61K 35/34**

(21) Application number: **89107552.5**

(22) Date of filing: **26.04.89**

(54) **Composition for repair of cartilage and bone and method for their preparation as skeletal tissue implant.**

(30) Priority: **29.04.88 US 187730**
05.12.88 US 280122

(43) Date of publication of application:
02.11.89 Bulletin 89/44

(45) Publication of the grant of the patent:
10.02.93 Bulletin 93/06

(84) Designated Contracting States:
AT BE CH DE ES FR GB GR IT LI LU NL SE

(56) References cited:
GB-A- 2 137 209

**RÖMPPS CHEMIE-LEXIKON, 8th ed.; pp. 253,
4325***

(73) Proprietor: **Itay, Samuel, Dr.**
16/3 Sharet Street
Kfar Saba(IL)

(72) Inventor: **Itay, Samuel, Dr.**
16/3 Sharet Street
Kfar Saba(IL)

(74) Representative: **Andrae, Steffen, Dr. et al**
Patentanwälte Andrae, Flach, Haug, Kneissl
Balanstrasse 55
W-8000 München 90(DE)

Note: Within nine months from the publication of the mention of the grant of the European patent, any person may give notice to the European Patent Office of opposition to the European patent granted. Notice of opposition shall be filed in a written reasoned statement. It shall not be deemed to have been filed until the opposition fee has been paid (Art. 99(1) European patent convention).

Description

Compositions for use in repairing bone and cartilage by implantation of material comprising a proliferating chondrocyte cell structure having phenotypic capability embedded in a vehicle or gel consisting of thrombin, antiprotease, fibrinogen, extracellular matrix and one or more growth factors that forms a "biological glue" of a biodegradable character are described in U.S. Patent No. 4,642,120. Before implantation, the cells are grown in tissue culture and harvested, and the chondrocyte population is embedded in the biological glue at a concentration of between from 100,000 to 500,000 cells per milliliter of glue. Before using this formulation, damaged cartilage and bone, as in a hip or other joint, are excised by surgery. A matching implant of the formulation is then inserted in the cavity, with or without bone segments to fill part of the volume. Cell proliferation continues in the permissive environment created by the system, while external influences are restricted. Chondrocytes (cartilage cells) and osteoblasts (bone forming cells) develop to unite with the existing structure, so that after a period of time, the implanted structure is virtually indistinguishable from the surrounding material.

Numerous advantages are derived from this approach in repair of articular cartilage, in comparison, for example, to replacement of a hip joint with a low friction metal-plastic prosthesis. The differences in the biomechanical properties between the bone and the prosthetic element is a major problem. Implantation of a prosthesis disables the mechanoreceptor system in the capsule of the joint which provides feedback for muscle control, resulting in wear and ultimately a need for replacement. The multiple freedoms of motion required of the joint, as for rotational and sliding movement, cannot be provided because of the absence of the mechanoreceptor system. In addition, the best low friction prosthetics have over 100 times the friction of the natural cartilage structure with the intervening synovial fluid, and for this reason also, wear and degradation are inevitable.

While the composition of Patent No. 4,642,120 has a demonstrated potential for repair of articular cartilage, it also has been recognized to have a number of limitations as a result of further experimental work. The needed cell proliferation capability was thought to be best available in embryonal chondrocytes (young committed chondrocytes) but for human use, availability is limited and major problems can arise from immune system reactions. Bone marrow stem cells are merely mentioned in the patent as a different possible source of cells, along with mesenchyme cells having potentiality for conversion to cartilage cells by self differentiation

or under the direction of chondrogenic factors. No work was done using these progenitors. Additional detailed information and discussion is contained in an article entitled "Use of Cultured Chick Epiphyseal Chondrocytes as Grafts for Defects in Chick Articular Cartilage", by S. Itay et al, Clinical Orthopedics, pp. 284-302, July, 1987.

The article mentioned cites a number of articles of general relevance to the topic as a whole. Three of these are of particular interest because they evidence attempts to transplant chondrocytes into articular cartilage that encountered limited success because, at least in part, of the absence of suitable biodegradable viscoelastic material and inability to produce cartilage. The three articles are: Bentley, G. et al, "Homotransplantation of isolated epiphyseal and articular cartilage chondrocytes onto joint surfaces of rabbits", Nature 230:385 (1971); Bentley, G. et al, "Isolated Epiphyseal chondrocyte allograft onto joint surface--An experimental study in rabbits", Ann. Rheum. Dis. 37:449 (1978); and Helbing, G. et al, "In vitro Untersuchungen an isolierten Chondrozyten zur Prognose von Knorpeltransplantaten", Helv. Chir. Acta 46:21 (1979).

As pointed out in U.S. Patent No. 4,642,120, it was previously thought that a limit had to be observed for chondrocyte concentrations of about 500,000 cells per milliliter of gel in order to avoid necrosis of the cells. Also, it was thought that only 5-50 units of thrombin per milliliter and about 25-80 mg/ml fibrinogen should be employed, with the setting of the gel being determined by the level of the thrombin, which should be kept at a limit of less than 50 units/ml. These relationships and parameters were found on further studies to limit proliferation rates and capacities, and capability for maturation and transformation of the implant into suitable phenotypic expressions, especially in large defects. Consequently, extension of this approach to repair of articular cartilage necessitates new compositions and procedures.

Compositions in accordance with the invention for regeneration of skeletal tissue employ cell cultures producing cells which express a chondrogenic phenotype. These include bone marrow derived chondrocytes and muscle fibroblast-derived chondrocytes as well as embryonal chondrocytes. Growth factors, mainly in the form of 10-20% serum, are employed in the culture medium to facilitate cell proliferation. A biological resorbable immobilization vehicle (BRIV) in the composition comprises preferably about 15-30% serum, 100-150 mg/ml of fibrinogen, 60-90 units/ml thrombin, 60 mM calcium chloride (CaCl_2), and 2000 units/ml (KIU) aprotinin. The cells in the implant are at a concentration of $80-160 \times 10^6$ cells/ml of BRIV. The resultant composition enables cells proliferation at

a higher rate *in vivo*, matures more quickly and transforms more readily into histological identity with surrounding cartilage and bone structures.

The present invention is directed to a composition for use as an implant in the repair of defects in cartilage and bone. The compositions are prepared by the isolation of the suitable cells of trypsinization and disruption of the tissue. The cells are cultivated on a suitable medium, harvested, and are combined with a fibrinogen-based biological resorbable immobilization vehicle (BRIV). The BRIV gel composition with embedded cells can be utilized immediately, can be stored for limited periods of times (2-3 days) in an incubator, cryopreserved for up to 3 years, or the harvested cells can be preserved for long periods of time by deep freezing and thawing just before use and then be embedded in the desired gel composition. When the implant is to be utilized, the damaged cartilage and bone are excised by surgery, the gel is immersed in a solution of fibrinogen, the site of implantation is sprayed with a thrombin solution, and the gel implant is pressed into the defect or injured site.

The cells utilized are preferably bone marrow progenital cells, but embryonal cells and chondrocytes derived from muscle fibroblast or other mesenchyme originated cells can also be used. The bone marrow progenital cells or the muscle fibroblast derived chondrocytes afford the advantage that the patient himself can be the donor at a convenient early time, with less chance of an auto immune response.

Culturing the cells on plastic provides an adequate growth environment for cell proliferation and for the selection of the desired cell type. The culturing medium contains 10-20% fetal calf serum, and may optionally contain growth factors such as IGF_I, IGF_{II}, TGF β , PDGF, or any other growth factors that will be found to facilitate the proliferation of the cells. The cells are subcultured until pure fibroblast-like cell populations are achieved. The pure fibroblast-like cell populations are trypsinized and placed in a suspension culture at a density of $3-8 \times 10^5$ cells/ml of medium and are cultured above soft agar in a F-12 medium (Sigma Co.) with 10% fetal calf serum (F.C.S.) and 50 μ g/ml of sodium ascorbate. After several days, the fibroblastic cells will be in aggregates of 30-60 cells.

Alternatively, the cells are trypsinized and the single cell suspension is cultured on Hma F-12 medium (Sigma Co.) on soft agar at a density of 2×10^5 to 4×10^5 cells/ml. 30-50 μ g/ml of sodium ascorbate is added daily to the medium.

The cells which are used in the implant are embedded in a biological resorbable immobilization vehicle (BRIV), a viscoelastic, biodegradable, biocompatible, resorbable matrix, at a concentra-

tion of $80-160 \times 10^5$ cells/ml of BRIV. The BRIV composition provides good adhesion and selective permeability of nutritive liquids and trophic agents. The BRIV gel also mechanically prevents cell migration. In addition, apart from fixing the chondrocytes in the defect sites, the BRIV also serves effectively as a proper extracellular milieu, supporting the growth and the differentiated state of the chondrocytes, while preventing fibroblast penetration and proliferation.

The composition of the BRIV comprises about 15-30% serum, 100-150 mg/ml of fibrinogen, 60-90 units/ml thrombin, 60mM calcium chloride (CaCl₂), and an antiprotease, such as aprotinin. Preferably, the BRIV implant composition comprises 20% fetal calf serum, 150 mg/ml fibrinogen, 90 units/ml of thrombin in 60mM of CaCl₂, and 2000 units/ml (KIU) aprotinin. The serum utilized in the BRIV can be fetal calf serum, umbilical cord serum from the second trimester, or horse serum, although fetal calf serum is preferred. Generally, a natural non-plasma protease inhibitor is used to prevent fast lysis of the matrix. A combination of polysaccharide inhibitors with plasma protease inhibitors and/or synthetic protease inhibitors can be used. Suitable protease inhibitors are chemical inhibitors such as epsilon-aminocaproic acid used in quantities of about 200 - 400 mg/ml of gel, and tranexemic acid, used in quantities of about 200-400 mg/ml of gel. Polysaccharide inhibitors can also be used. Also, a natural non-plasma protease inhibitor such as anti-trypsin (Chicken egg white, Sigma, type III) may be used or suitable synthetic protease inhibitors. Plasma protease inhibitors may also be utilized.

The bioimplants of the invention can be utilized in various ways. A freshly prepared implant (BRIV composition containing embedded cells) can be applied directly to the injured site or defect. In addition, the BRIV composition with embedded cells can be covered with F-12 medium (Sigma Company) plus 10% fetal calf serum and can be stored in a CO₂ incubator for a 2-3 days, at about 37°C. Further, an implant can be prepared from cryopreserved cells (90% fetal calf serum and 10% DMSO) in liquid nitrogen (cryopreserved for up to 3 years) which have been thawed and then embedded in BRIV. As a preferable option, the cells can be collected by centrifugation, and the complete implant with cells embedded in BRIV can be cryopreserved (90% fetal calf serum and 10% dimethyl sulfoxide (DMSO)) in liquid nitrogen for longer periods. The implant in this case is thawed in the operating room and immediately implanted. An implant may optionally contain bone segments or bone substitute to fill part of the volume.

When the implant is to be utilized, the injured site or defect is sprayed with a thrombin solution, and the BRIV matrix containing cells is immersed

in a solution containing fibrinogen and antiprotease. The implant is pressed into the injured site or defect so as to fill up the defect. The thrombin solution comprises 90 units/ml thrombin in 60mM CaCl_2 .

Within 48 hours after implantation, chondrocyte proliferation is seen. Two weeks later, hyaline cartilage matrix surrounds these cells. Within eight weeks, the defects are completely filled with hyaline cartilage which integrates smoothly with the neighboring cartilage without the formation of fibrous tissue at the interface. The cell content and the rate of proteoglycan synthesis in the reparative tissue remains high for four months, then declines slowly towards the level of the surrounding cartilage. Six months after transplantation, the cartilaginous repair tissue at the level below the ossification front shows penetration by vascular elements and cartilage to bone transformation. Eighteen months after transplantation, all the implant below the ossification front transforms into bony elements, while the articular part of the implant remains cartilaginous with all the properties of the original cartilage. No signs of immunogenic rejection or degenerative changes of the implant are observed. On the contrary, the articular surface of the implanted area appears to be younger than the original surface.

The composition of the invention permits cell proliferation at a higher rate *in vivo*. The cells in the implant mature more quickly, and transform more readily into histological identity with the surrounding cartilage and bone structures. Within two months of transplantation, the defect is filled properly with active proliferating cells, and is integrated well with no fibrocartilage or other soft tissue at the edges. At 2 to 6 months, all the implant that is below the osteochondral junction is transformed into bone while articular cartilage retains its cartilaginous properties. The biodegradable BRIV of the implant has been reabsorbed.

The following specific Examples can be used to further illustrate the invention which contain a best mode. The Examples were prepared and tested as described.

Example 1

Implant Containing Bone Marrow Derived Chondrocytes

Autologous or homologous bone marrow is obtained by aspiration with a bone biopsy needle from the iliac crest or femoral canal. The aspirated cells are injected into phosphate buffered saline (PBS) containing 0.25% trypsin and injected sequentially through 17, 18 and 20 gauge needles to achieve a single cell suspension. Higher gauge

needles are found to induce some cell destruction. The cells are plated at a density of $50\text{--}100 \times 10^6$ cells on 100 mm tissue culture dishes fed with BGJb medium (GIBCO) with 15% F.C.S. The medium is changed daily or as required by the proliferation rate of the cells. The medium may be supplemented by growth factors such as IGF_I, IGF_{II}, TGF β , PDGF or any other growth factors that will be found to facilitate the proliferation of the cells. The cells are subcultured weekly and after 5-6 subculturings, an almost pure fibroblastic stromal cell population is achieved. This cell population is then trypsinized and put in a suspension culture at a density of $3\text{--}8 \times 10^6$ cells/ml of medium and cultured above soft agar in a F-12 medium (Sigma Co.) with 10% F.C.S. and 50 $\mu\text{g/ml}$ sodium ascorbate added daily to the medium. The fibroblastic stromal cells start to aggregate immediately and after three-seven days, most of the cells are in aggregates of 30-60 cells. All the aggregates express a chondrogenic phenotype, as determined by employing histochemical and immunohistochemical probes for analysis.

Although bone marrow derived chondrocytes are preferred in this example, one can use chondrocytes or osteoblasts of autologous or homologous origin, or homologous committed chondrocytes, or any other progenital cells of mesenchymal origin. It can be seen that this initial formulation comprises purification, proliferation and manipulation of a population expressing a chondrogenic or osteogenic phenotype. More specifically, the proliferating cells are from the class comprising bone marrow stroma cells, embryonal committed chondrocytes and any undifferentiated mesenchymal cells.

To incorporate the cells in a biodegradable viscoelastic matrix, the resulting pellet of cells is resuspended in a small volume of phosphate buffer saline (PBS) containing fibrinogen (150 mg/ml) and 20% of fetal calf serum and aprotinin, available under the trademark "Trasylol®" (2000 KIU/ml) or another antiprotease. The solution contains cells (ranging in concentration between $80\text{--}160 \times 10^6$ cells/ml), fibrinogen, 15-30% serum and antiprotease and may be designated solution A. Specifically in this example 120×10^6 cells/ml of BRIV, 20% fetal calf serum, 150 mg/ml fibrinogen, 90 units/ml of thrombin in 60 mM CaCl_2 , and 2,000 units of aprotinin are employed. A second solution, designated as solution B, comprises thrombin (90 units/ml in 60 mM CaCl_2). The solutions are mixed, keeping the ratio of solutions A and B 3:1 (v/v). The implant is immersed in F-12 medium containing 10% F.C.S. and may be immediately used. Alternatively, the implant may be cryopreserved (in LN_2 , for example) in 90% F.C.S. and 10% DMSO (or any other cryopreservation regime). At trans-

plantation, the defect is sprayed with a thrombin solution and the implant is press fitted into the defect.

Data collected in experimentation with bone marrow derived chondrocytes and embryonal derived chondrocytes in several species (avian and mammalian) by macroscopic observation, histological sections, and biochemical test showed that at the site of transplantation within two months the defect is filled properly with a complete congruency at the articular surface and perfect integration with no fibrocartilage or other soft tissue at the interfaces. At 2 to 6 months all the implant that is below the osteochondral junction is transformed into bone while articular cartilage retains its cartilaginous properties. No degenerative changes or immunological rejection is observed after prolonged follow-up periods.

Although the serum is preferentially fetal calf serum in this example, umbilical cord serum from the second trimester or horse serum or any combination of these may be employed. No extracellular matrix need be used.

Example 2

Preparation of Composition for Cartilage Repair

As starting material, epiphysis of long bones (tibia, femur, humerus) was used. The isolation procedure of embryonal chondrocytes comprises trypsinization of the epiphysis (1% porcine trypsin), incubation for 60 minutes at 37°C. and vortexing for 2 minutes in each 10 minute interval and thereafter a gentle mechanical disintegration of the tissue by a Teflon channeled homogenizer. Trypsin activity is terminated by serum which contains an antiproteolytic substance. The resulting single cell suspension is then seeded for several days (4-7 days) in Ham F-12 medium (Sigma Co.) on plates coated with soft agar (0.5% Bacto-agar in Ham F-12) at a density of 2×10^5 to 4×10^5 cells/ml. An amount of 50 µg of sodium ascorbate is added daily to the medium. During this growth period, most of the fibroblasts are dying off and chondrocyte enrichment does occur. The cells are collected by centrifugation and used directly in the BRIV as a fresh graft. Alternatively, the complete graft may be cryopreserved, or the cells may be cryopreserved (90% fetal calf serum (F.C.S.) and 10% dimethyl sulfoxide (DMSO)) in liquid nitrogen for longer periods and embedded at a later date in BRIV. The cells were collected and embedded at a concentration of $80-160 \times 10^6$ cells/ml of the same viscoelastic biodegradable matrix as in Example 1. The results obtained are comparable to those of Example 1.

Example 3

Implant Containing Muscle Fibroblast-Derived Chondrocytes

Autologous or homologous muscle is obtained by an open biopsy under local anesthesia. The muscle is minced into very small pieces of tissue and is then trypsinized for 30 minutes in 0.5% trypsin in P.B.S. and Ethylene Diamine Tetraacetic Acid (E.D.T.A.) with occasional vortexing. The trypsinized cells are then filtered through a 53 micron Nitex filter. The trypsinization is then stopped by M.E.M. (minimal essential) medium (GIBCO Co.) containing 15% F.C.S. After centrifugation, the pellet of cells is washed with M.E.M. medium containing 15% F.C.S. The cells are then plated at a density of $50-100 \times 10^6$ cell per 100 mm tissue culture dishes and are maintained daily with M.E.M. medium with 15% F.C.S. The cells are subcultured weekly on M.E.M. medium with 15% F.C.S. and after three subculturings, a pure fibroblastic-like population is achieved. This cell population is then trypsinized and put in a suspension culture at a concentration of $3-8 \times 10^6$ cells/ml. of medium and cultured above soft agar in F-12 medium (Sigma Co.) with 10% F.C.S. and 50 µg/ml of sodium ascorbate added daily to the medium. The fibroblastic cells start to aggregate immediately and after three to seven days most of the cells are in aggregates of 30-60 cells. All the aggregates express a chondrogenic phenotype as determined by employing histochemical and immunohistochemical probes for analysis.

Although muscle-derived chondrocytes are preferred in this example, one can use bone marrow-derived chondrocytes or committed chondrocytes as well. The cells are collected by centrifugation and directly embedded at a concentration of $80-160 \times 10^6$ cells/ml of the same viscoelastic biodegradable matrix (BRIV) as in Example 1. Alternatively, the cells can be cryopreserved (in 90% F.C.S. and 10% DMSO) in liquid nitrogen for long periods, thawed before usage, embedded in BRIV at a concentration of $80-160 \times 10^6$ cells/ml and used. As a further alternative, the cells can be embedded in BRIV, with the entire implant being cryopreserved in 90% F.C.S. and 10% DMSO, and thawed before usage in the operating room. The results obtained are comparable to those of Example 1.

Example 4

Preparation of Composition for Bone Repair

In order to regenerate bone defects, one of three methods may be used.

4A. For small defects 2-4 cm in length, one uses

an implant as proposed in Example 1, Example 2, or Example 3.

4B. For large defects, a composition graft of bone substitute used as a supporting matrix with biomechanical properties near to the properties of a native bone is used. The cells are combined with this matrix via the biodegradable fibrinogen based adhesive matrix.

4C. The bone marrow stromal cells can be induced in vitro to express an osteoblastic phenotype and used directly as in 4A or 4B to correct bony defects. (This can be used only in the autologous group where the bone marrow originates from the patient with the bone defect.)

The Process

A traumatic (fracture) or pathologic (tumor) or degenerative disease defect in bone or articular cartilage is cleaned up and shaped into geometric configuration (cuboidal or cylindrical). In the case of an articular surface, the entire procedure can be done through an arthroscopic device.

After the damage area is prepared, a frozen implant with an identical shape (prepared as described in detail in Example 1, Example 2 or Example 3) is rapidly thawed by putting it into saline at 37°C. for 5-10 minutes.

The implant is then immersed in a solution of fibrinogen and the implantation site is sprayed with the thrombin solution. The implant is now pressed into the defect. In an articular defect, continuous passive motion is started immediately.

In the case of large defects, a composite graft of the biological implant embedded in (or above) a bone substitute material of suitable shape can be used. This implant will be either custom made or as a commercial standard type.

While various alternatives and modifications are proposed above, it will be appreciated that the invention is not limited thereto but encompasses all forms and variations in accordance with the appended claims.

Claims

1. A composition for the regeneration of articular cartilage and bones by implantation comprising bone marrow derived chondrocytes or osteoblasts of autologous or homologous origin, or homologous committed chondrocytes, or autologous or homologous muscle fibroblast derived chondrocytes or any other progenital cells from mesenchymal origin and a biodegradable, biocompatible, biological resorbable immobilization vehicle (BRIV) adhesive glue consisting of fibrinogen, thrombin, CaCl_2 , protease inhibitor, and about 10-30% serum, characterized in that said composition contains from $80-160 \times 10^6$ cells/ml of BRIV.

2. The composition according to claim 1, wherein said serum comprises 15-30% serum selected from fetal calf serum or umbilical cord serum from the second trimester or horse serum or any combination of such serums.

3. The composition according to claim 1, wherein the cells are of bone marrow derived chondrocytes at a concentration of 120×10^6 cells/ml of BRIV that can be induced in culture to proliferate and be converted to express chondrogenic or osteogenic phenotypes.

4. The composition according to claim 1, wherein the cells are of muscle fibroblasts derived chondrocytes at a concentration of 120×10^6 cells/ml of BRIV that can be induced in culture to proliferate and be converted to express chondrogenic or osteogenic phenotypes.

5. The composition of claim 1, wherein the protease inhibitor is selected from the group consisting of polysaccharide inhibitors, plasma protease inhibitors, and synthetic and natural non-plasma protease inhibitors.

6. A method of preparing a skeletal tissue implant comprising a composition in accordance with any of the preceding claims, which methods comprises the steps of:

- purification, proliferation and manipulation of a cell population expressing a specific chondrogenic or osteogenic phenotype; and
- embedding the cells in a biological resorbable immobilization vehicle (BRIV) comprising at least 10%, preferably 15-30%, serum so that the concentration of the cells is between 80×10^6 and 160×10^6 cells/ml of BRIV.

7. The method according to claim 6, wherein the proliferating cells are selected from bone marrow stroma cells, embryonal committed chondrocytes, or chondrocytes derived from muscle fibroblasts and any undifferentiated mesenchymal cells.

8. The method according to claim 6 or 7, wherein the BRIV includes 100-150 mg/ml fibrinogen and 60-90 units/ml thrombin, and wherein the BRIV further includes about 2,000 units/ml aprotinin.

9. The method according to any of claims 6 to 8,

wherein the cells are embedded in a BRIV in a suitably shaped bone substitute.

10. The method according to any of claims 6 to 9, comprising the steps of:

a. culturing a proliferating cell population comprising autologous bone marrow stroma cells or autologous muscle fibroblast derived chondrocytes;
b. manipulating them in suspension above soft agar at a concentration in excess of 2×10^6 cells/ml of medium for at least five days;
c. harvesting the cells; and
d. embedding the cells at a concentration in excess of 80×10^6 cells/ml of BRIV comprising in excess of 10% serum, 100 mg/ml fibrinogen, and 60 units/ml thrombin in an excess of 40 mM CaCl_2 and 2000 units aprotinin.

11. The method according to claim 10, wherein the BRIV includes 15-30% serum and 60 units/ml thrombin in an excess of 60 mM CaCl_2 .

12. The method according to claim 10, wherein the biological implant is stored under low temperature conditions after preparation with a preservative medium containing 90% fetal calf serum, and 10% DMSO, and wherein there is about 20% serum.

13. The method according to any of claims 10 to 12, wherein the harvested cells are stored under low temperature conditions after preparation with a preservative medium containing 90% fetal calf serum and 10% DMSO, and embedded at a later date in BRIV.

14. The method according to claim 10, wherein the serum is selected from fetal calf serum, umbilical cord serum from the second trimester, and horse serum.

Patentansprüche

1. Eine Zusammensetzung zur Regenerierung von Gelenkknorpel und Knochen durch Implantation, die knochenmark-stämmige Chondrozyten oder Osteoblasten von autologem oder homologem Ursprung, oder homologe festgelegte Chondrozyten, oder autologe oder homologe muskelfibroblasten-stämmige Chondrozyten oder irgendwelche andere Vorläuferzellen von mesenchymalem Ursprung und einen biologisch abbaubaren, biokompatiblen, biologisch resorbierbaren Immobilisationsvehikel-(BRIV)-Haftkleber, der aus Fibrinogen, Thrombin,

CaCl_2 , Proteaseinhibitor und etwa 10-30% Serum besteht, umfaßt, dadurch gekennzeichnet, daß die genannte Kombination von 8 bis 160×10^6 Zellen/ml BRIV enthält.

2. Zusammensetzung nach Anspruch 1, bei der das genannte Serum 15 bis 30% Serum darstellt, das ausgewählt ist aus fötalem Kälberserum oder Nabelschnurserum aus dem zweiten Trimester oder Pferdeserum oder irgendeiner Kombination derartiger Sera.

3. Zusammensetzung nach Anspruch 1, bei der die Zellen knochenmark-stämmige Chondrozyten in einer Konzentration von 120×10^6 Zellen/ml BRIV sind, die in Kultur zur Vermehrung angeregt werden können und so umgewandelt werden können, daß sie chondrogene oder osteogene Phänotypen hervorbringen.

4. Zusammensetzung nach Anspruch 1, bei dem die Zellen muskelfibroblasten-stämmige Chondrozyten in einer Konzentration von 120×10^6 Zellen/ml BRIV sind, die in Kultur zur Vermehrung angeregt werden können und die so umgewandelt werden können, daß sie chondrogene und osteogene Phänotypen hervorbringen.

5. Zusammensetzung nach Anspruch 1, bei der der Proteaseinhibitor aus der Gruppe ausgewählt ist, die aus Polysaccharidinhibitoren, Plasmaproteaseinhibitoren und syntetischen und natürlichen Nichtplasmaproteaseinhibitoren besteht.

6. Verfahren zur Herstellung eines Skelettgewebeimplantats, das eine Zusammensetzung gemäß irgendeinem der vorausgehenden Ansprüche umfaßt, wobei das Verfahren die Schritte umfaßt:

a. Reinigung, Vermehrung und Manipulation einer Zellpopulation, die einen spezifischen chondrogenen oder osteogenen Phänotyp hervorbringt; und;
b. Einbetten der Zellen in ein biologisch resorbierbares Immobilisationsvehikel (BRIV), das wenigstens 10%, vorzugsweise 15 bis 30% Serum aufweist, so daß die Konzentration der Zellen zwischen 80×10^6 und 160×10^6 Zellen/ml BRIV liegt.

7. Verfahren nach Anspruch 6, bei dem die sich vermehrenden Zellen ausgewählt sind aus Knochenmark-Stromazellen, embryonalen festgelegten Chondrozyten, oder Chondrozyten, die muskelfibroblasten-stämmig sind, sowie irgendwelchen undifferenzierten mesenchymalen Zellen.

8. Verfahren nach Anspruch 6 oder 7, bei dem das BRIV 100 bis 150 mg/ml Fibrinogen und 60 bis 90 Einheiten/ml Thrombin enthält und bei dem das BRIV außerdem etwa 2000 Einheiten/ml Aprotinin enthält. 5
9. Verfahren nach irgendeinem der Ansprüche 6 bis 8, bei dem die Zellen in einem BRIV in einem geeignet geformten Knochenersatz eingebettet sind. 10
10. Verfahren nach irgendeinem der Ansprüche 6 bis 9, das die Schritte umfaßt:
 - a. Kultivieren einer sich vermehrenden Zellpopulation, die autologe Knochenmark-Stromazellen oder autologe muskelfibroblastenstämmige Chondrozyten umfaßt; 15
 - b. deren Manipulation in Suspension über weichem Agar in einer Konzentration von mehr als 2×10^6 Zellen/ml Medium für wenigstens 5 Tage; 20
 - c. Ernten der Zellen; und
 - d. Einbetten der Zellen in einer Konzentration von mehr als 80×10^6 Zellen/ml BRIV, das mehr als 10 % Serum, 100 mg/ml Fibrinogen und 60 Einheiten/ml Thrombin in einem Überschuß von 40mM CaCl_2 und 2000 Einheiten Aprotinin umfaßt. 25
11. Verfahren nach Anspruch 10, bei dem das BRIV 15 bis 30% Serum und 60 Einheiten/ml Thrombin in einem Überschuß von 60mM CaCl_2 enthält. 30
12. Verfahren nach Anspruch 10, bei dem das biologische Implantat nach seiner Herstellung unter Niedertemperaturbedingungen mit einem Konservierungsmedium gelagert wird, das 90% fötales Kälberserum und 10% DMSO enthält, und wobei etwa 20% Serum vorliegen. 35 40
13. Verfahren nach irgendeinem der Ansprüche 10 bis 12, bei dem die geernteten Zellen nach ihrer Herstellung unter Niedertemperaturbedingungen mit einem Konservierungsmedium gelagert werden, das 90% fötales Kälberserum und 10% DMSO enthält, und bei dem sie zu einem späteren Zeitpunkt in BRIV eingebettet werden. 45
14. Verfahren nach Anspruch 10, bei dem das Serum ausgewählt ist aus fötalem Kälberserum, Nabelschnurserum aus dem zweiten Trimester und Pferdeserum. 50

Revendications

1. Composition pour la régénération de cartilage

articulaire et d'os par implantation comprenant des chondrocytes ou des ostéoblastes provenant de moelle osseuse d'origine autologue ou homologue, ou des chondrocytes différenciés homologues, ou des chondrocytes provenant de fibroblastes musculaires autologues ou homologues ou toutes autres cellules progénitales d'origine mésenchymateuse et une colle adhésive formant un véhicule d'immobilisation résorbable biologique (BRIV), biocompatible, biodégradable constituée de fibrinogène, de thrombine, de CaCl_2 , d'inhibiteur de protéase et d'environ 10-30 % de sérum, caractérisée en ce que ladite composition contient de $80\text{--}160 \times 10^6$ cellules/ml de BRIV.

2. Composition suivant la revendication 1, caractérisée en ce que le sérum comprend 15-30 % de sérum choisi parmi du sérum de veau foetal ou du sérum de cordon ombilical du second trimestre ou du sérum de cheval ou l'une quelconque combinaison de ces sérums.
3. Composition suivant la revendication 1, caractérisée en ce que les cellules sont constituée de chondrocytes provenant de moelle osseuse à une concentration de 120×10^6 cellules/ml de BRIV qui peuvent être induites en culture pour proliférer et être converties pour exprimer des phénotypes chondrogéniques ou ostéogéniques.
4. Composition suivant la revendication 1, caractérisée en ce que les cellules sont des chondrocytes provenant de fibroblastes musculaires à une concentration de 120×10^6 cellules/ml de BRIV qui peuvent être induites en culture pour proliférer et être converties pour exprimer des phénotypes chondrogéniques ou ostéogéniques.
5. Composition suivant la revendication 1, caractérisée en ce que l'inhibiteur de protéase est choisi dans le groupe comprenant les inhibiteurs de polysaccharide, les inhibiteurs de protéase plasmatiques et les inhibiteurs de protéase non plasmatiques synthétiques et naturels.
6. Procédé de préparation d'un implant de tissu du squelette comprenant une composition suivant l'une quelconque des revendications précédentes, lequel procédé comprend les étapes suivantes :
 - a. la purification, la prolifération et la manipulation d'une population cellulaire exprimant un phénotype chondrogénique ou ostéogénique spécifique, et

b. l'incorporation des cellules dans un véhicule d'immobilisation résorbable biologique (BRIV) comprenant au moins 10 %, de préférence 15-30 % de sérum de telle sorte que la concentration des cellules se situe entre 80×10^6 et 160×10^6 cellules/ml de BRIV.

7. Procédé suivant la revendication 6, caractérisé en ce que les cellules proliférantes sont choisies parmi des cellules de stroma de moelle osseuse, des chondrocytes différenciés embryonnaires ou des chondrocytes provenant de fibroblastes musculaires et toutes cellules mésenchymateuses non différenciées. 5 10
8. Procédé suivant l'une ou l'autre des revendications 6 et 7, caractérisé en ce que le BRIV comprend 100-100 mg/ml de fibrinogène et 60-90 unités/ml de thrombine et en ce que le BRIV comprend de plus environ 2000 unités/ml d'aprotinine. 15 20
9. Procédé suivant l'une quelconque des revendications 6 à 8, caractérisé en ce que les cellules sont incorporées dans un BRIV dans un produit de remplacement osseux façonné de façon appropriée. 25
10. Procédé suivant l'une quelconque des revendications 6 à 9, comprenant les étapes suivantes 30
 - a. la culture d'une population cellulaire proliférante comprenant des cellules de stroma de moelle osseuse autologues ou des chondrocytes provenant de fibroblastes musculaires autologues, 35
 - b. leur manipulation en suspension au-dessus d'agar mou à une concentration excédant 2×10^6 cellules/ml de milieu pendant au moins cinq jours; 40
 - c. la récolte des cellules, et
 - d. l'incorporation des cellules à une concentration excédant 80×10^6 cellules/ml de BRIV comprenant plus de 10 % de sérum, 100 mg/ml de fibrinogène et 60 unités/ml de thrombine dans un excès de 40 mM de CaCl_2 et 2000 unités d'aprotinine. 45
11. Procédé suivant la revendication 10, caractérisé en ce que le BRIV comprend 15-30 % de sérum et 60 unités/ml de thrombine dans un excès de 60 mM de CaCl_2 . 50
12. Procédé suivant la revendication 10, caractérisé en ce que l'implant biologique est conservé sous des conditions de basse température après la préparation avec un milieu de conser-

vation contenant 90 % de sérum de veau foetal et 10 % de DMSO et en ce qu'il y a environ 20 % de sérum.

13. Procédé suivant l'une quelconque des revendications 10 à 12, caractérisé en ce que les cellules recueillies sont conservées sous des conditions de basse température après la préparation avec un milieu de conservation contenant 90 % de sérum de veau foetal et 10 % de DMSO et en ce qu'elles sont incorporées à une date ultérieure dans du BRIV. 5 10
14. Procédé suivant la revendication 10, caractérisé en ce que le sérum est choisi parmi du sérum de veau foetal, du sérum de cordon ombilical du second trimestre et du sérum de cheval. 15 20